



**PRESS RELEASE (2025/07/04)**

## Uncovering the mechanism behind dual-end cleavage in transfer RNAs

Advancing synthetic biology and biotechnology, a study reveals how a 12-unit enzyme complex precisely cuts both ends of transfer RNA

Fukuoka, Japan—To build proteins, cells rely on a molecule called transfer RNA, or tRNA. tRNAs act like protein-building couriers, where they read the genetic instructions from messenger RNA, mRNA, and deliver the right amino acids to ribosomes, the cell's protein-making factories. But before tRNAs can do their work, they first need to be trimmed and shaped properly.

Now, researchers from Kyushu University have revealed that the smallest known protein-based tRNA-processing enzyme, called HARP, forms a star-shaped complex of 12 protein molecules, making it capable of cutting both the 5' and 3' ends of tRNA. The team hopes that their findings will have a broad impact on synthetic biology and biotechnology research, and aid in the designing of artificial enzymes and RNA processing tools. Their findings were published in the journal [\*Nature Communications\*](#).

In any biological system, most proteins that are made in the cell need to undergo processing for them to fully work. In the case of tRNA, one of those processes is the cutting of the straggling ends of the RNA that make up the molecule. Depending on the direction, these are called 5-prime or 3-prime ends, denoted as 5' and 3', respectively.

One key enzyme responsible for cutting the extra segment at the 5' end of the tRNA is RNase P. Found in almost all life forms, this enzyme exists in two broad forms: one that is mostly made of RNA and another that is entirely protein-based. The RNA-based version usually forms a large, complex structure with several proteins and has been well studied over the past 40 years.

On the other hand, protein-only RNase P enzymes are more streamlined. These come in two main types: PRORP, which is found in higher organisms like plants and animals, and HARP, which is found in certain bacteria and archaea. HARP—short for Homologs of Aquifex RNase P36—is known for its small size and unique six-pointed, star-like structure. But how it performs such a complex task—or why it forms such a distinctive shape—remained unclear.

"To investigate and visualize HARP bound to pre-tRNA and uncover how it processes the molecule, we used cryogenic electron microscopy (cryo-EM) single-particle analysis," explains [Professor Yoshimitsu Kakuta](#) from [Kyushu University's Faculty of Agriculture](#), who led the study.

The researchers found that the overall structure of the enzyme with the pre-tRNAs had a radial structure with pre-tRNA molecules alternately bound to five binding sites on the enzyme. Cryo-EM analysis showed that the 12-subunit HARP enzyme acts like a "molecular ruler," measuring the distance from the 5' end to the "elbow" of the pre-tRNA to precisely identify the cleavage site. Remarkably, this mechanism was also observed in other types of RNase P enzymes, indicating a case of convergent evolution across different organisms.

"Our structural analysis shed light on how HARP processes the 5' leader sequence and revealed that the functional 12-subunit HARP complex binds only five pre-tRNA molecules, not ten as previously predicted," adds [Assistant Professor Takamasa Teramoto](#), the first author of the study. "This means that 7 of the enzyme's 12 active sites remain unoccupied."

When the team conducted cleavage assays to understand the functionality of these vacant sites, they found a second cleavage product that corresponded to the 3' end of the pre-tRNA. This was a new finding. It suggests that HARPs first trim the extra nucleotides at the 5' end and then use the remaining empty active sites to carry out the cleavage at the 3' end.

"The oligomerization of the small protein HARP confers it with bifunctionality in pre-tRNA processing. Our findings illustrate an evolutionary strategy by which organisms with compact genomes can acquire multifunctionality," concludes Kakuta.

Uncovering such evolutionary strategies where limited structural elements are arranged flexibly to gain new functions could assist in the development of future tools in synthetic biology and biotechnology.

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For more information about this research, see "Structural basis of transfer RNA processing by bacterial minimal RNase P," Takamasa Teramoto, Takeshi Koyasu, Takashi Yokogawa, Naruhiko Adachi, Kouta Mayanagi, Takahiro Nakamura, Toshiya Senda, Yoshimitsu Kakuta, Nature Communications, <https://doi.org/10.1038/s41467-025-60002-1>

### **About Kyushu University**

Founded in 1911, [Kyushu University](#) is one of Japan's leading research-oriented institutes of higher education, consistently ranking as one of the top ten Japanese universities in the Times Higher Education World University Rankings and the QS World Rankings. The university is one of the seven national universities in Japan, located in Fukuoka, on the island of Kyushu—the most southwestern of Japan's four main islands with a population and land size slightly larger than Belgium. Kyushu U's multiple campuses—home to around 19,000 students and 8000 faculty and staff—are located around Fukuoka City, a coastal metropolis that is frequently ranked among the world's most livable cities and historically known as Japan's gateway to Asia. Through its [VISION 2030](#), Kyushu U will "drive social change with integrative knowledge." By fusing the spectrum of knowledge, from the humanities and arts to engineering and medical sciences, Kyushu U will strengthen its research in the key areas of decarbonization, medicine and health, and environment and food, to tackle society's most pressing issues.

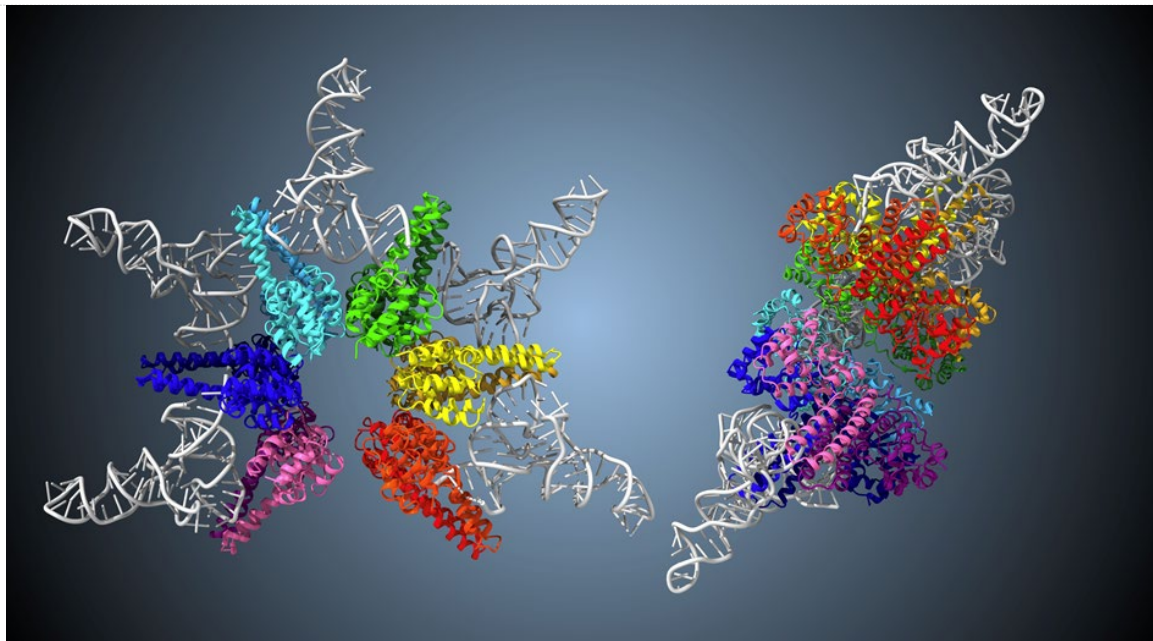


Fig. 1. Star-shaped small enzyme with “dual-functionality” helps in proper tRNA functioning. HARPs are the smallest known RNase P enzymes that help with tRNA maturation in bacteria and archaea. How such a small enzyme accomplishes this crucial task has baffled researchers for some time. Now, researchers from Kyushu University, Japan, have revealed that HARPs form a 12-subunit star-shaped structure with binding sites to process both the ends (5' and 3') of pre-tRNAs. This novel observation, where small enzymes take up different forms to perform multiple functions, suggests molecular evolution of these crucial enzymes. (Takamasa Teramoto/Yoshimitsu Kakuta/Kyushu University)

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